Root Necrosis of Strawberry Caused by Colletotrichum *acutatum*¹

J. C. Mertely, B. B. Forcelini, and N. A. Peres²

Colletotrichum acutatum is widely known as a fruit rot pathogen, but it also infects other strawberry organs, including the roots. Root necrosis has been observed in Florida since 2000.

Pathogens and Symptoms

Transplants with infected root systems often grow poorly or fail to become established after overhead irrigation is withdrawn.Few functional roots are found on infected plants even 1 to 2 weeks after transplant (Figure 1). Old structural roots are brown or black with few feeder roots, whereas new roots develop brown lesions, die back from the tip, or fail to emerge from the crown (Figure 2). In severe cases, *C. acutatum* enters the crown, causing a basal crown rot and eventually killing the plant (Figure 3). Surviving plants are stunted or variable in size, flower late, and produce a poor early crop (Figure 4). Infected plants may recover during the cool winter months and produce normally in February and March, if an outbreak of anthracnose fruit rot does not occur.

Disease Development and Spread

C. acutatum frequently colonizes leaves and petioles of runner plants in the nursery. Symptoms may not be visible in the nursery environment, but if inoculum is allowed to build up and the weather is favorable, flowers and fruit are attacked and lesions may develop on the petioles (Figure 5).

Little is known about how or when the pathogen spreads from colonized tissue above the ground to the root system below. However, *C. acutatum* grows freely in diseased tissues and has been isolated from the soil around diseased plants. Healthy plants are presumably contaminated by this inoculum during normal digging, trimming, and packing operations in the nursery.



Figure 1. Root necrosis symptoms (few functional roots). Credits: UF/IFAS GCREC

- 1. This document is PP-211, one of a series of the Plant Pathology Department, UF/IFAS Extension. Original publication date August 2005. Revised January 2009, September 2012, August 2017, and February 2022. Visit the EDIS website at https://edis.ifas.ufl.edu for the currently supported version of this publication.
- 2. J. C. Mertely, former biological scientist IV; B. B. Forcelini, former graduate assistant; and N. A. Peres, professor, Plant Pathology Department; UF/IFAS Gulf Coast Research and Education Center, Wimauma, FL.

The Institute of Food and Agricultural Sciences (IFAS) is an Equal Opportunity Institution authorized to provide research, educational information and other services only to individuals and institutions that function with non-discrimination with respect to race, creed, color, religion, age, disability, sex, sexual orientation, marital status, national origin, political opinions or affiliations. For more information on obtaining other UF/IFAS Extension publications, contact your county's UF/IFAS Extension office. U.S. Department of Agriculture, UF/IFAS Extension Service, University of Florida, IFAS, Florida A & M University Cooperative Extension Program, and Boards of County Commissioners Cooperating. Andra Johnson, dean for UF/IFAS Extension.



Figure 2. Root necrosis symptoms (root lesions). Credits: UF/IFAS GCREC



Figure 3. Basal crown rot caused by *Colletotrichum acutatum*. Credits: UF/IFAS GCREC

Early in the season, disease spread below ground is unlikely since the root systems are relatively isolated; however, above-ground spread in the foliage does occur and may be facilitated by overhead irrigation during establishment. Observations made at the UF/IFAS GCREC suggest that disease severity is influenced by the degree of plant stress during establishment. Even cultivars that are only moderately susceptible to anthracnose fruit rot, e.g., 'Florida Radiance' and 'Sensation[®] Florida 127', are susceptible to root necrosis disease.



Figure 4. Stunted plants due to root necrosis. Credits: UF/IFAS GCREC

Control

Diseases caused by *C. acutatum* are best controlled by exclusion (not introducing the pathogen into the field). Therefore, transplants should be purchased from a reputable source. However, this does not guarantee disease-free material. For that reason, transplants should be inspected for petiole lesions caused by *C. acutatum* (Figure 5).



Figure 5. Petiole lesions caused by *Colletotrichum acutatum*. Credits: UF/IFAS GCREC

When the pathogen is present and susceptible cultivars are being grown, pre-plant fungicide dips may be used to suppress disease development. Abound[®], Oxidate[®], Actinovate[®], and Switch[®] are currently labeled for this use (Table 1). These products have been tested at UF/IFAS GCREC—Balm by dipping naturally infected runner plants for 5 minutes just before planting. Plants dipped in Switch survived better and produced higher marketable yields in all trials. Actinovate has worked well in some trials but not others and Oxidate was not effective in our trials. Abound has performed well in our trials until the 2013-2014 season, when lack of ARN control was reported in FL production areas even after transplants were dipped in Abound (Forcelini et al. 2016). Control failure was due to the emergence of C. acutatum populations resistant to the strobilurin fungicides. Growers should consider dip treatments mainly for cultivars that are known to be highly susceptible and/ or when transplants are from nurseries with a history of C. acutatum.

Measures that reduce plant stress during establishment may also reduce the severity of root necrosis disease. Whenever possible, strawberries should be planted in the morning to avoid high temperatures and drying conditions that occur in the afternoon. After setting, overhead irrigation should be started as soon as possible to prevent transplants from wilting on the hot plastic. After 7 to 8 days, plant response should be used to determine if overhead irrigation is still necessary. Irrigation should be continued in the afternoon if drying conditions are encountered or the plants wilt when water is withdrawn. If hot weather is expected after overhead irrigation is withdrawn, consider spraying the beds with Surround[®], which reduces heat stress by coating the plants and plastic with white kaolin clay.

Literature Cited

Forcelini, B. B., Seijo, T. E., Amiri, A., and Peres, N. A. 2016. "Resistance in strawberry isolates of *Colletotrichum acutatum* from Florida to quinone-outside inhibitor fungicides". *Plant Dis.* 100:2050–2056.

Table 1. Products labeled in Florida for management of root necrosis.

Trade Name	Active ingredient	Туре	REI* (hours)	Comments
Abound	azoxystrobin	strobilurin	4	Pre-plant dip in 5 to 8 fl oz of 2.08 lb flowable formulation per 100 gal water.
Actinovate	Streptomyces lydicus	biological	1	See label for specific rates and application recommendations.
Oxidate	hydrogen peroxide	oxidizer disinfectant	**	Pre-plant dip in 64 fl oz of 27% formulation per 100 gal water. Higher rates (e.g., 128 fl oz) may cause stunting or phytotoxicity.
Switch	cyprodinil + fludioxonil	pyrimidine + pyrrole	12	Pre-plant dip in 5 to 8 oz of 62.5WG formulation in 100 gal water. One-year plant back restriction.
Surround	kaolin clay	stress reducer	4	Add 25 to 50 lbs to 100 gal water and spray over tops of beds. Gradually washes off plants and plastic during subsequent rains.

* PHI = Post harvest interval. REI = Restricted entry interval.

** After the application has dried. Recommendations: Follow label instructions closely when dip treating transplants. To avoid possible phytotoxicity, carefully monitor treatment times, and set the transplants as soon as possible after dipping. Recommendations given in this fact sheet are based on experimentation and statements from the manufacturer. Consult your product label for specific use requirements and restrictions.